

Isolation and Structure Elucidation of Deformylflustrabromine from the North Sea Bryozoan *Flustra foliacea*

Nicola Lysek^a, Eike Rachor^b and Thomas Lindel^{c,*}

^a Pharmazeutisch-chemisches Institut der Universität, Im Neuenheimer Feld 364, D-69120 Heidelberg, Germany

^b Alfred-Wegener-Institut für Polar- und Meeresforschung, Columbusstraße, D-27568 Bremerhaven

^c Department Chemie der Universität, Butenandtstr. 5–13, D-81377 München, Germany. Fax: (+49)89/21 80-77 34. E-mail: thomas.lindel@cup.uni-muenchen.de

* Author for correspondence and reprint requests

Z. Naturforsch. **57c**, 1056–1061 (2002); received February 6, 2002

Bromindole, Bryozoan, Inverse Prenylation

The brominated pyrrolo[2,3-b]indole deformylflustrabromine was isolated as a new natural product from the bryozoan *Flustra foliacea*, collected in the North Sea. Deformylflustrabromine appears to be the missing link in the biosynthetic sequence from flustrabromine to flustraminol A. Flustramines A, D, and dihydroflustramine C were determined as other major constituents of the investigated sample. Deformylflustrabromine is cytotoxic against the human colon cancer cell line HCT-116 (IC₅₀ 5.8 µM).

Introduction

The bryozoan *Flustra foliacea* (Flustridae) is the source of unique brominated pyrrolo[2,3-b]indoles sharing their condensed heterocyclic system with the potent acetylcholine esterase inhibitor physostigmine (**1**; Fig. 1), a plant secondary metabolite being a candidate for the treatment of Alzheimer's disease (Witkop, 1998). More than 15 tryptophan-

derived alkaloids have been so far isolated from *Flustra* (Holst *et al.*, 1994, and references cited therein). Searching for possible biosynthetic intermediates of the *Flustra* secondary metabolites we re-investigated the organism. In a specimen collected in the southern North Sea, we detected a new monobrominated secondary metabolite with a relative mass of 320/322 which may represent the missing link towards flustraminol A (**8**, Fig. 3; Carlé and Christophersen, 1981).

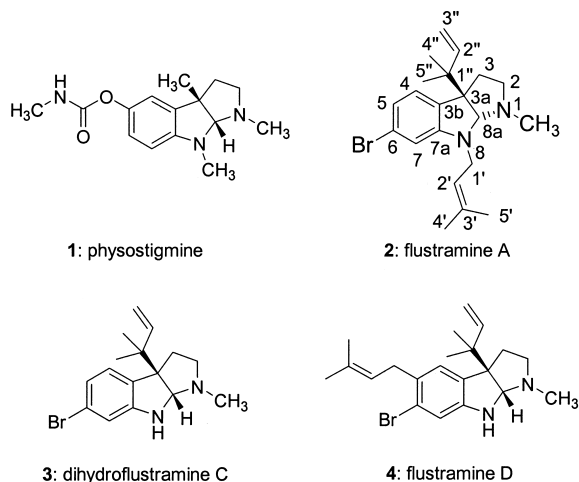


Fig. 1. The potent acetylcholine esterase inhibitor physostigmine (**1**) and three known bromindole alkaloids (**2**, **3**, **4**) re-isolated from the bryozoan *Flustra foliacea*.

Experimental Section

General

Column chromatography was carried out on Sephadex LH-20 (Pharmacia) and on silica gel (particle size 230–400 mesh, Merck). Thin-layer chromatography (TLC) was performed on silica gel (precoated silica gel plate F₂₅₄ Merck). Preparative HPLC separation columns (25 × 250 mm) were prefilled with LiChroprep RP-8 (25–40 µm, Merck) or LiChroprep Si-60 (25–40 µm, Merck). The peaks were detected at 254 nm. NMR spectra were recorded on Bruker WM 250, WM 300, AM 360 or Varian INOVA-400 spectrometers. The NMR shifts were calibrated using the NMR solvent as internal reference. All infrared spectra were recorded on a Perkin Elmer 1600 series

FT-IR spectrometer. The UV/Vis-spectra were recorded using a Hewlett-Packard UV-spectrometer HP 8452 Diode Array System. Fast atom bombardment (FAB) mass spectra were recorded on a JEOL JMS-700 mass spectrometer with nitrobenzyl alcohol as matrix. The optical rotation was recorded using a Perkin-Elmer 241 A polarimeter with a 10 cm cell.

Collection

The material was collected with an otter trawl from the sea floor along the margin of the deep Helgoland trench in the southeastern North Sea at about 54 °12' N and 7 °47' E and a water depth of 33 m–45 m during the expedition no. 116 of RV “Heincke” on 12 February 1999. When the haul was on board, *Flustra foliacea* was selected from the catch, washed with sea water and deep-frozen in a PE bag.

Isolation

The bryozoan *Flustra foliacea* (82.5 g, fresh weight) was lyophilized and extracted with MeOH/CH₂Cl₂ (1:1 v/v, 750 ml, 3 times). After concentration the crude extract (13.9 g) was partitioned between isooctane and MeOH. The MeOH phase was washed with isooctane (3 times) and concentrated. The residue was partitioned between *n*-BuOH and water. The *n*-BuOH phase was washed with water and the residue (4.08 g) was fractioned by gel chromatography (Sephadex LH-20, MeOH).

Flustramine A (2)

Obtained from fraction 4 of the Sephadex LH-20 column via flash chromatography (silica, gradient CHCl₃ to CHCl₃/MeOH 70:30 v/v). 16 mg (0.02% of the dry weight); ¹H NMR (250 MHz, [D₁]chloroform): δ = 6.90 (d, *J* = 8.5 Hz, 1H, 4-H), 6.69 (dd, *J* = 1.5, 8.5 Hz, 1H, 5-H), 6.48 (d, *J* = 1.5 Hz, 1H, 7-H), 5.94 (dd, *J* = 10.5, 17.0 Hz, 1H, 2''-H), 5.22 (t, *J* = 6.3 Hz, 1H, 2'-H), 5.06 (dd, *J* = 1.5, 10.5 Hz, 1H, 3''-H_E), 4.98 (dd, *J* = 1.5, 17 Hz, 1H, 3''-H_E), 4.35 (s, 1H, 8a-H), 3.84 (d, *J* = 6.3 Hz, 2H, 1'-H), 2.67 (m, 2H, 2-H), 2.42 (s, 3H, NCH₃), 2.22 (m, 2H, 3-H), 1.73 (s, 6H, 4'-H, 5'-H), 1.01 (s, 3H, 4''-H), 0.94 (s, 3H, 5''-H); ¹³C NMR (90.6 MHz, [D₁]chloroform): δ = 153.6 (C-7a), 144.9 (C-2''),

134.6 (C-3'), 132.6 (C-3b), 125.8 (C-4), 121.7 (C-6), 120.9 (C-2'), 119.1 (C-5), 113.1 (C-3''), 109.3 (C-7), 89.3 (C-8a), 63.4 (C-3a), 53.2 (C-2), 45.9 (C-1'), 41.3 (C-1''), 37.8 (NCH₃), 34.4 (C-3), 25.6 (C-4'), 23.5 (C-4''), 22.3 (C-5''), 18.1 (C-5'); IR (NaCl): $\tilde{\nu}$ = 2965, 2972, 2855, 1674, 1594, 1487 cm⁻¹; UV (EtOH): λ_{max} (ϵ) = 214 (18248), 262 (6626), 316 nm (2645 mol⁻¹ · cm⁻¹ · l); [α]_D²⁰ = -76.92 (*c* = 6.6 mm in EtOH); FABMS *m/z* (%) = 389/391 (49/40) [M⁺], 319/321 (100/98); HRFABMS calcd. for C₂₁H₃₀⁸¹BrN₂ 391.1572, found 391.1579.

Deformylflustrabromine (5)

Fraction 5 of the Sephadex column was purified by HPLC (flow 11.5 ml · min⁻¹; RP-8, H₂O/MeOH/HOAc (50:50:0.1 v/v/v, *t_R* 10.06 min)); then (flow 11.5 ml · min⁻¹; silica, gradient CHCl₃ to CHCl₃/CH₃OH (70:30, over 30 min, *t_R* 25.00 min)). 59 mg (0.072% of the dry weight); ¹H NMR (360 MHz, [D₁]chloroform/[D₄]methanol (7:3 v/v)): δ = 7.53 (s, 1H, 7-H), 7.45 (d, *J* = 8.3 Hz, 1H, 4-H), 7.15 (d, *J* = 8.3 Hz, 1H, 5-H), 6.15 (dd, *J* = 17.3, 10.4 Hz, 1H, 2''-H), 5.17 (d, *J* = 17.3 Hz, 1H, 3''-H_E), 5.16 (d, *J* = 10.4 Hz, 1H, 3''-H_Z), 3.23 (m, 2H, 1'-H), 3.08 (m, 2H, 2'-H), 2.71 (s, 3H, NCH₃), 1.53 (s, 6H, 4'', 5''-H); ¹³C NMR (90.6 MHz, [D₆]DMSO): δ = 145.5 (C-2''), 141.8 (C-2), 135.5 (C-7a), 127.6 (C-3a), 121.2 (C-5), 119.4 (C-4), 113.5 (C-6), 113.4 (C-7), 111.6 (C-3''), 104.9 (C-3), 48.5 (C-2'), 45.4 (NCH₃), 38.7 (C-1''), 27.6 (C-4'', C-5''), 21.6 (C-1'); IR (NaCl): $\tilde{\nu}$ = 3279, 2978, 2760, 2433, 1710, 1591, 1466 cm⁻¹; UV (MeOH): λ_{max} (ϵ) = 204 (30190), 230 (22731), 282 nm (4768 mol⁻¹ · cm⁻¹ · l); FABMS *m/z* (rel. intensity): 321/323 (100/96) [M⁺]; HRFABMS calcd. for C₁₆H₂₂⁷⁹BrN₂ 321.0966, found 321.0974.

Results and Discussion

The purification protocol stepwise applied solvent partitioning, gel gravity chromatography (Sephadex LH-20) and preparative HPLC using RP-8 and silica stationary phases. For a graphical representation of the isolation procedure see Fig. 2. The molecular formula of the new compound **5** was determined as C₁₆H₂₂BrN₂ by HRFABMS. On the basis of connectivity information derived from HSQC, HMBC, and COSY NMR experiments, the computer program Cocon (Lindel *et al.*, 1999; Köck *et al.*, 1999) calculated

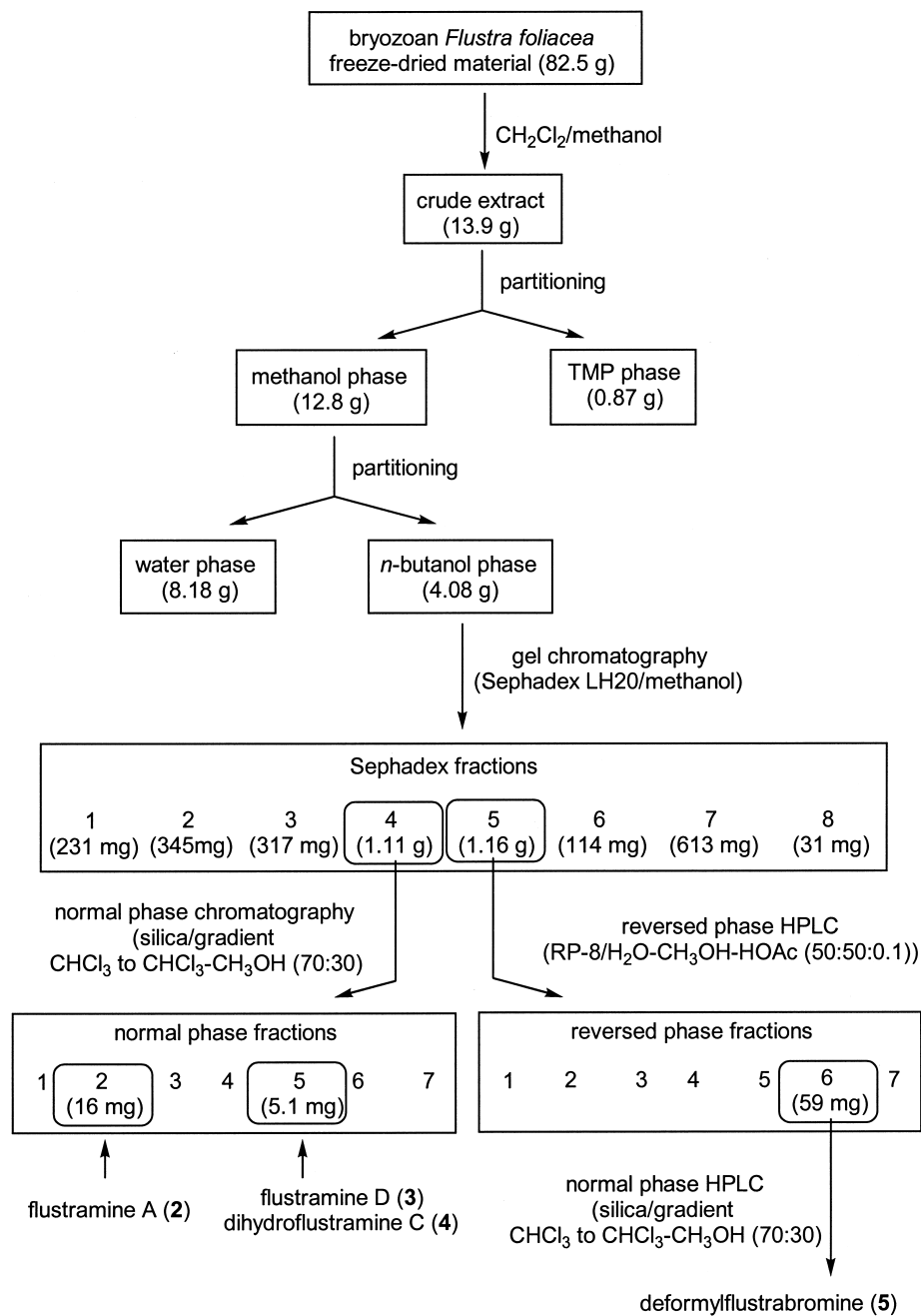


Fig. 2. Isolation protocol leading to the new natural product deformylflustrabromine (5) and the known metabolites flustramine A (2), dihydroflustramine C (3) and flustramine D (4).

two constitutions fulfilling all required constraints. Due to the absence of experimental HMBC correlations of the methyl protons in [D₆]DMSO (see Table I), an alternative constitution was generated in which the bromo and methylamino substituents have changed places. An increment calculation for

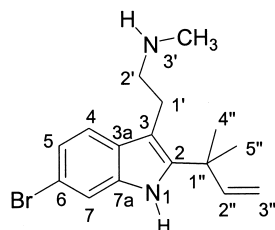
C-6 of the indole ring and biosynthetic considerations strongly favor constitution 5. The ¹H-NMR-spectrum of 5 shows the patterns of a 6-substituted indole. In addition to the new compound 5, the known brominated pyrrolo[2,3-b]indoles flustramine A (2; Carlé and Christophersen, 1979, 1980),

dihydroflustramine C (**3**; Wright, 1984; Laycock *et al.*, 1986), and flustramine D (**4**; Laycock *et al.*, 1986) were isolated. Very recently, the isolation of **5** was independently described by Peters *et al.* (2002).

Given the close relationship of the new compound **5** to flustrabromine (**7**; Wulff *et al.*, 1981), **5** should be named deformylflustrabromine. Deformylflustrabromine (**5**) is no artefact, because the ^1H -NMR-signals of this major secondary metabolite were clearly visible in the *n*-butanol phase of the investigated specimen of *Flustra foliacea*, before any step of chromatography. There was no signal of any *N*-formyl proton (to be expected at about δ 8.0 in the ^1H -NMR-spectrum) present at any stage of the extraction. The content of deformylflustrabromine (0.072% of the dry weight) compares to that of other constituents, in particular of flustrabromine (**7**; Wulff *et al.*, 1981, 1982), 6-bromoformyltryptamine (**6**; Wulff *et al.*, 1982), and flustramine A (**2**; *e.g.*, 0.035% of the dry weight, Carlé and Christophersen, 1980).

The inverse prenylation of the 2-position of indole alkaloids is under intense investigation (for a review, see Williams *et al.*, 2000). From the lack of stereospecificity observed in the construction of the quaternary center at the indole 2-position, Stocking *et al.* (1999, 2000) concluded that the olefinic π -system of dimethylallylpyrophosphate is introduced by a “reverse” prenyl transferase presenting both faces of the π -system to the 2-position of the indole. In all of the investigated cases, the α -amino function of the tryptophan unit was acylated, *e.g.*, as a diketopiperazine.

In the secondary metabolism of *Flustra*, a formyl group may play the analogous role. Flustrabromine (**7**; isolated as a mixture of *E*-/*Z*-isomers) would be formed from 6-bromo-*N*₆-methyl-*N*₆-formyltryptamine (**6**) via inverse prenylation. Both **6** and **7** have been isolated as natural products from *Flustra foliacea* (Wulff *et al.*, 1982). In the continued sequence from flustrabromine (**7**) to the natural product flustraminol A (**8**; Carlé and Christophersen, 1981), the new metabolite defor-



5: deformylflustrabromine

Table I. NMR spectroscopical data of deformylflustrabromine (**5**).

Position	^1H NMR ^a	^{13}C NMR ^b	HMBC correlations ^c
2	—	141.8	
3	—	104.9	
3a	—	127.6	
4	7.45 (1H, d, 8.3 Hz)	119.4	3, 3a, 6, 7a
5	7.15 (1H, d, 8.3 Hz)	121.2	7
6	—	113.5	
7	7.53 (1H, s)	113.4	3a, 5, 6
7a	—	135.5	
1'	3.23 (2H, m)	21.6	2, 3, 3a
2'	3.08 (2H, m)	48.5	
1''	—	38.7	
2''	6.15; dd; 17.3, 10.4	145.5	2, 1'', 4''(5'')
3''	5.17 (1H, d, 17.3 Hz; <i>E</i>); 5.16 (1H, d, 10.4 Hz; <i>Z</i>)	111.6	1'', 2''
4''	1.53 (3H, s)	27.6	2, 1'', 2'', 5''
5''	1.53 (3H, s)	27.6	2, 1'', 2'', 4''
NCH ₃	2.71 (3H, s)	45.4	

^a 360 MHz ([D₁]chloroform/
[D₄]methanol (7:3)). ^b 90.6 MHz
([D₆]DMSO). ^c Position numbers are
given.

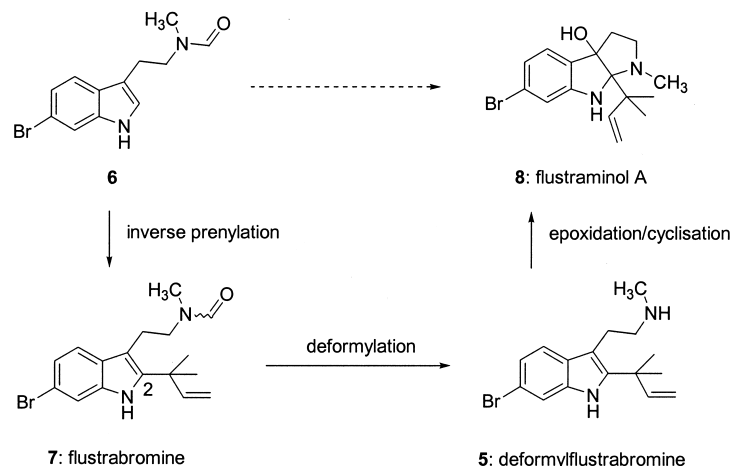


Fig. 3. Deformylflustrabromine (**5**) as a possible missing link in the biosynthesis of flustraminol A (**8**), starting from the natural product **6**.

mylflustrabromine (**5**) may be the missing link. Epoxidation of **5**, followed by ring opening of the epoxide ring would lead to flustraminol A (**8**), as outlined in Fig. 3. Indeed, a biomimetic cyclisation was recently induced by treatment of *N*-Boc-tryptophan methyl ester, inversely prenylated at the 2-position of the indole ring, with dimethyldioxirane (Schkeryantz *et al.*, 1999). The stereochemistry of the natural product **8** was not determined.

While the crude extract of *Flustra foliacea* was not cytotoxic, the purified compounds showed activity against the human colon cancer cell line HCT-116. The new natural product deformylflustrabromine (**5**) showed the highest, but still

moderate, cytotoxicity of $1.87 \mu\text{g} \cdot \text{ml}^{-1}$ ($5.8 \mu\text{M}$, IC_{50} , HCT-116). Flustramine A (**2**), D (**4**), and dihydroflustramine C (**3**) were weakly cytotoxic in the range of $10 \mu\text{g} \cdot \text{ml}^{-1}$ ($26 \mu\text{M}$, IC_{50}).

Acknowledgements

This work was funded in part by the Bundesministerium für Bildung und Forschung (BMBF grant V-258, cooperation with the BASF AG, Ludwigshafen). We are grateful to Tanja Mülhaupt and Prof. Dr. William H. Fenical (Scripps Institution of Oceanography, San Diego, USA) for performing the cytotoxicity assays.

- Carlé J. S. and Christophersen C. (1979), Bromo-substituted physostigmine alkaloids from a marine bryozoa *Flustra foliacea*. *J. Am. Chem. Soc.* **101**, 4012–4013.
- Carlé J. S. and Christophersen C. (1980), Marine alkaloids. 2. Bromo alkaloids from the marine bryozoan *Flustra foliacea*. Isolation and structure elucidation. *J. Org. Chem.* **45**, 1586–1589.
- Carlé J. S. and Christophersen C. (1981), Marine alkaloids. 3. Bromo-substituted alkaloids from the marine bryozoan *Flustra foliacea*, Flustramine C and Flustraminol A and B. *J. Org. Chem.* **46**, 3440–3443.
- Holst P. B., Anthoni U., Christophersen C. and Nielsen P. H. (1994), Marine alkaloids, 15. Two alkaloids, flustramine E and debromoflustramine B, from the marine bryozoan *Flustra foliacea*. *J. Nat. Prod.* **57**, 997–1000.
- Köck M., Junker J., Maier W., Will M. and Lindel T. (1999), A Cocon analysis of proton-poor heterocycles-application of carbon chemical shift predictions for the evaluation of structural proposals. *Eur. J. Org. Chem.* 579–586.
- Laycock M. V., Wright J. L. C., Findlay J. A. and Patil A. D. (1986), New physostigmine related bromoalkaloids from the marine bryozoan *Flustra foliacea*. *Can. J. Chem.* **64**, 1312–1316.
- Lindel T., Junker J. and Köck M. (1999), 2D NMR-guided constitutional analysis of organic compounds employing the computer program Cocon. *Eur. J. Org. Chem.* 573–577.
- Peters L., König G. M., Terlau H. and Wright A. D. (2002), Four New Bromotryptamine Derivatives from the Marine Bryozoan *Flustra foliacea*. *J. Nat. Prod.* **65**, web release date September 11, 2002.
- Schkeryantz J. M., Woo J. C. G., Siliphaivanh P., Depew K. M. and Danishefsky S. J. (1999), Total synthesis of gypsetin, deoxybrevianamide E, brevianamide E, and tryprostatin B: Novel constructions of 2,3-disubstituted indoles. *J. Am. Chem. Soc.* **121**, 11964–11975.
- Stocking E. M., Williams R. M. and Sanz-Cervera J. F. (1999), Reverse und “normale” Prenyltransferasen haben unterschiedliche Seitenselektivitäten bei der Biosynthese von Paraherquamid. *Angew. Chem.* **111**, 880–883; *Angew. Chem. Int. Ed.* **38**, 786–789.
- Stocking E. M., Williams R. M. and Sanz-Cervera J. F. (2000), Reverse prenyl transferases exhibit poor facial discrimination in the biosynthesis of paraherquamide A, brevianamide A, and austamide. *J. Am. Chem. Soc.* **122**, 9089–9098.
- Williams R. M., Stocking, E. M. and Sanz-Cervera, J. F. (2000), Biosynthesis of prenylated alkaloids derived from tryptophan. *Top. Curr. Chem.* **209**, 97–173.
- Witkop B. (1998), From the “Ordeal bean” (*Physostigma venenosum*) to the ordeal of Alzheimer’s disease – Some of the legacy of Percy Lavon Julian (1899–1975). *Heterocycles* **49**, 9–27.
- Wright J. L. C. (1984), A new antibiotic from the marine bryozoan *Flustra foliacea*. *J. Nat. Prod.* **47**, 893–895.
- Wulff P., Carlé J. S. and Christophersen C. (1981), Marine alkaloids. Part 4. A Formamide, flustrabromine, from the marine bryozoan *Flustra foliacea*. *J. Chem. Soc. Perkin Trans. 1*, 2895–2898.
- Wulff P., Carlé J. S. and Christophersen C. (1982), Marine alkaloids – 5. Flustramide A and 6-bromo-*N*₆-methyl-*N*₆-formyltryptamine from the marine bryozoan *Flustra foliacea*. *Comp. Biochem. Physiol.* **71B**, 523–524.